

Biological Forum – An International Journal

13(4): 45-50(2021)

ISSN No. (Print): 0975-1130 ISSN No. (Online): 2249-3239

Management of Early Blight of Tomato through Neem Formulation and Bio-inoculants

Anand Choudhary*, J.R. Verma, Dama Ram and Pooja Yadav Department of Plant Pathology, CoA, Jodhpur (Agriculture University, Jodhpur- 342304, (Rajasthan), India.

> (Corresponding author: Anand Choudhary*) (Received 02 August 2021, Accepted 04 October, 2021) (Published by Research Trend, Website: www.researchtrend.net)

ABSTRACT: Early blight disease is painful nerve to tomato growing farmer because it causes huge economic loss to the farmer every year. *Alternaria solani* cause early blight of tomato and it is considered weed of field because of its wide adaptability under different environment. Use of fungicides for the management of disease in crop puts a large number of negative health and environmental effects therefore, the urgent need for a more sustainable and ecological approach to manage disease without fungicides. To avoid relying solely on chemicals and to identify a viable alternative component, five neem formulations and four bio-inoculants were evaluated by Poison Food Technique and Dual Culture Technique in the lab in the year of 2020-21 against *A. solani*. Among neem formulations, neem oil (71.91%) was found effective inhibiting mycelial growth of *A. solani* followed by multineem (58.49%), Neem excel (48.92%) and Repellent plus (38.00%) at the mean of two concentration (1000 & 1500 ppm). In case of bio-inoculants, *T. harzianum* was found most effective inhibiting mycelial growth 71.15% followed by *T. viride* (66.35%).

Keywords: Alternaria solani, Early blight, Neem Formulation, Bio-inoculants, Mycelial inhibition.

INTRODUCTION

The Portuguese introduced the tomato (*Solanum lycopersicum* L) to India in the 1700s (Kale and Kale, 1994). Tomato is also known as "Poor Man's Orange" in India because it is a nutrient-dense super food and an excellent source of antioxidants, which help to reduce the risk of heart disease and cancer. As a result, tomato fruits are in high demand throughout the year. The tomato is the world's fourth most cultivated crop, with a production of 130 million tons and an area of 5.2 million hectares (Anonymous, 2020). It is an inevitable vegetable crop world over and of course, for India.

India is the world's second largest tomato producer after China, with an area of 778 thousand hectares and a production of 19397 metric tons, accounting for 11% of global production (Anonymous, 2020). Cultivated tomatoes have a narrow genetic diversity as a result of intense selection and inbreeding during evolution and domestication, these species are more susceptible to disease epidemics during the growing season (Zhang *et al.*, 2002).

Early blight disease is caused by several species of *Alternaria* including *Alternaria solani*, *A. alternata* as well as *A. tomatophila* (Adhikari *et al.*, 2017). Worldwide, early blight disease is one of the dreadful diseases of tomato resulting up to 78% yield and production loss (Datar and Mayee 1981; Bessadat *et al.*, 2014). It directly harms the plant and reduces both the quantity and quality of the economic yield. It has a significant impact on crop growth at all stages during

both *Kharif* and *Rabi* season. This disease, which can cause severe defoliation in severe condition, is most damaging to tomato in areas with heavy rainfall, high humidity and fairly high temperatures 24-29°C (Peralta *et al.*, 2005). Epidemics can occur in semi arid climates where frequent and prolonged nightly dews occur (Vennila *et al.*, 2020).

Currently, the control of plant disease, pest in only through by spaying a large amount of synthetic fungicides (Cook, 2000). However, an increase use of synthetic fungicide can severely deteriorate the planet's health (Wang *et al.*, 2010). Different types of fungicides have been used for the control of Alternaria blight, but fungicide treatment is not economically feasible, nor environmentally sound. Fungicides are first applied 1-2 days after transplantation and then require routine application at the interval of 7 to10 days for effective control, thereby increasing production cost and environment pollution (Kemmitt 2002).

Neem oil has been the cure for many fungal diseases caused by Alternaria solani, Aspergillus flavus, Alternaria alternate (Girish and Shankara, 2008). Different neem formulation founds antifungal properties against Alternaria spp. (Saha et al., 2005; Guleria and Kumar, 2006). Kota, (2003) has reported that bioagents T. harzianum and T. virens were highly inhibiting mycelial growth of A. alternata in vitro. Kumar, (2008) evaluated effect of four bio-agents Trichoderma spp., namely Trichoderma harzianum, T. konigii, T. viride and T. virens, against growth of A. alternata under laboratory conditions. In his

Choudhary et al.,

experiments, *T. harzianum* has shown highest inhibitory effect on radial growth (RG) rate of *A. alternata*. *Trichoderma spp.* have ability to detoxify pesticides and herbicide have been revealed in several findings (Vazquez *et al.*, 2015; Zafra *et al.*, 2015).

The effects of systematic fungicides are inactivated by fungi enzymes (Golyshin, 1990). Under mutation impact, fungicides lose their inhibitory effect. Therefore, we urgent need to identify other viable alternative component to manage disease of plants. Keeping in view the importance of sustainable and ecological approach to manage disease, the present study was conducted to assess the *in vitro* efficacy of different neem formulation and bio-agents against *Alternaria solani*.

MATERIAL AND METHODS

Evaluation of neem formulations under in *in vitro* **condition.** A total five neem formulations (Table 1) along with control were used in present study in the year of 2020-21. Bio efficacy of different neem formulations were tested *in vitro* by "Poisoned Food Technique". The poisoned food technique was used for study of neem formulation at 1000 ppm and 1500 ppm

concentration was mixed with potato dextrose agar medium. The experiment was conducted in the CRD with three replications. Percent inhibition of mycelial growth over untreated control was calculated by applying the formula given by Vincent, (1947).

Poison Food Technique. Required quantity of each neem formulations under study was mixed thoroughly in sterilized 100 ml PDA media filled in 250 ml flask separately under aseptic condition. The medium was supplemented with streptomycin sulphate @ 50 ppm to prevent bacterial contamination. The poisoned medium was then poured in sterilized petri plates (20 ml) and allowed it to solidify. Mycelium discs of 5 mm size from seven days old culture was cut by a sterile cork borer and one such disc was placed at the center of each agar plate. The plate without any neem formulations served as control. Three replications were maintained for each concentration. Such plates were incubated at room temperature and the radial growth was measured when fungus attained maximum growth in control plates. Percent inhibition of mycelial growth over untreated control was calculated by applying the formula given by Vincent, (1947).

Table 1: List of Neem formulations evaluated against A. solani by Dual Culture Technique (in vitro).

Sr. No.	Treatment	Doses	s in ppm
1.	Multineem	1000	1500
2.	Neemix	1000	1500
3.	Repellent Plus	1000	1500
4.	Neem oil	1000	1500
5.	Neem Excel	1000	1500
6.	Control		

In vitro evaluation of bio-inoculants. *In vitro* evaluation was carried out with two fungal and two bacterial bio-agents (Table 2) along with control against *A. solani* by Dual Culture Technique (Denis and Webster, 1971) in the year of 2021-21. Both bio-agents and test fungus were cultured on potato dextrose agar media.

Dual Culture Technique. Twenty ml of sterilized and cooled potato dextrose agar was poured into sterile petri plates and allowed to solidify. For evaluation of fungal bio-agents, mycelial discs of test fungus were inoculated at one end of the petri plate and antagonistic fungus was placed opposite to it on the other end.

In case of bacterial bio-agents evaluation, the bacteria were streaked one day earlier at one end of the petri plate or at the middle of the petri plate and the test fungus placed at the opposite end. The plates were incubated at $27\pm1^{\circ}$ C and zone of inhibition was recorded by measuring the clear distance between the margin of the test fungus and antagonistic organism. The colony diameter of pathogen in control plate was also recorded. The colony diameter of the pathogen in control plates will be also recorded.

The experiment will be conducted in the CRD with four replications. Percent inhibition of mycelial growth over untreated control was calculated by applying the formula given by Vincent, (1947).

Table 2: List of bio-agents evaluated against *A. solani* by Dual Culture Technique (*in vitro*).

Sr. No.	Bio-agent	
1.	Trichoderma viride	
2.	Trichoderma harzianum	
3.	Pseudomonas fluorescens	
4.	Bacillus subtillis	
5	Control	

RESULTS AND DISCUSSION

Management of early blight of tomato through neem formulations under *in vitro* condition. Five neem formulations with two different concentrations (1000 & 1500 ppm) along with control were evaluated *in vitro* against *A. solani* by applying Poisoned Food Technique (Nene and Thapliyal, 1993). The observations on per cent growth inhibition (PGI) were recorded and the results were presented in Table 3 Fig. 1, Plate 1a &1b.

Sr. No.		Percent mycelium growth inhibition*		
	Treatment	Concentration (ppm))
		1000	1500	Mean
1.	Multineem	52.19 (46.24)	64.79 (53.58)	58.49
2.	Neemix	36.04 (36.87)	46.15 (42.77)	41.09
3.	Repellent plus	34.15 (35.74)	41.85 (40.29)	38.00
4.	Neem oil	66.73 (54.79)	77.09 (61.39)	71.91
5.	Neem excel	40.96 (39.77)	56.87 (48.93)	48.92
6.	Control	0.00	0.00	0.00
0.	Colluor	(0.00)	(0.00)	
Factor		S. Em ±	$n \pm CD (p = 0.05)$	
Neem formulation (N)		0.460	1.349	
Concentration (C)		0.265	0.779	
Interaction (N x C)		0.650	1.908	

Table 3: Efficacy of neem formulations against Alternaria solani in vitro condition.

*Average of three replications

Figures in parentheses are angular transformed values

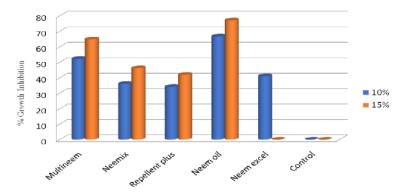
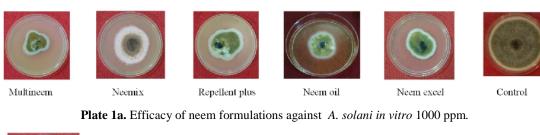


Fig. 1. Effect of neem formulations on mycelial growth inhibition (%) of A. solani in vitro.



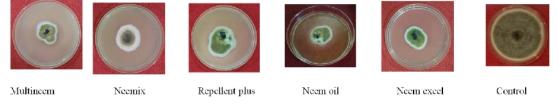


Plate 1b. Efficacy of neem formulations against A. solani in vitro 1500 ppm

The result presented data in Table 3 showed that neem oil (77.09 %) at 1500 ppm was found best and significantly superior over all other neem formulation concentrations in inhibiting mycelial growth of *A. solani* followed by multineem (64.79 %) at 1500 ppm concentrations. Neem oil at 1000 ppm concentrations was more effective as compare to multineem at 1500 ppm concentrations. Least mycelial inhibition was observed in repellent plus (34.15%) at 1000 ppm concentration. In case of neem excel, it inhibiting 56.87% mycelial growth of pathogen at 1500 ppm

concentrations. Least inhibition i.e. 34.15% & 41.85% was observed in repellent at both concentrations, respectively. Since, no precise information was available on the efficacy of neem formulation against *A. solani* caused early blight in tomato.

Management of early blight of tomato through bioinoculants in *in vitro* conditions. Four antagonists' *viz.*, *Trichoderma viride*, *T. harzianum*, *Pseudomonas fluorescence* and *Bacillus subtilis* (Plate 2) were studied under laboratory condition for their antagonism against *Alternaria solani by* dual culture technique.

Choudhary et al.,

Biological Forum – An International Journal 13(4): 45-50(2021)

The results revealed that all the antagonists significantly helped in inhibiting the mycelial growth of *A. solani* over control. Significantly highest per cent mycelial growth inhibition was observed in *T harzianum* (71.15%) followed by *T. viride* (66.35%)

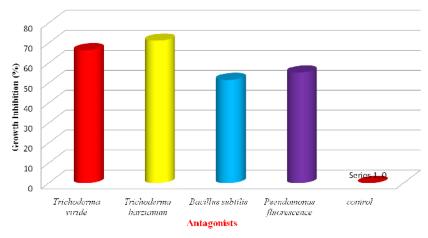
and *Pseudomonas fluorescence* (55.25%) after 7 day of incubation. While bio-inoculants *Bacillus subtilis* showed minimum per cent mycelial growth inhibition (51.57%) (Table 4, Fig. 2 and Plate 2).

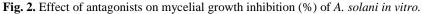
Table 4: Effect of	different	bio-inoculants	against A.	solani in vitro.

Sr. No.	Bio-inoculants	Per cent inhibition of mycelial growth*
1.	Trichoderma viride	66.35 (54.52)
2.	Trichoderma harzianum	71.15 (57.49)
3.	Bacillus subtilis	51.57 (45.88)
4.	Pseudomonas fluorescence	55.25 (47.99)
5.	Control	0.00 (0.00)
	S.Em ±	0.354
	CD (p = 0.05)	1.718

*Average of four replications

Figures in parentheses are angular transformed values





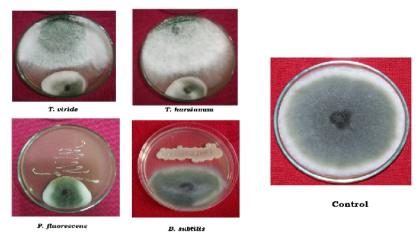


Plate 2. Bio-efficacy of Bio-inoculants against A. solani in vitro.

It is evident from the above results that neem leaf formulation protected from early blight pathogen (*Alternaria solani*) of the tomato plants. *In vitro* tests carried out by Chaudhary *et al.*, (2003) reported that extracts of *A. indica* gave the second highest inhibition of *A. alternata* (54%). Neem formulation derived from the Neem tree (*Azadirachta indica*) have been found to be fetal against a broad spectrum of plant pathogenic fungi such as *Alternaria solani*, *Fusarium oxysporum* (Bhonde *et al.*, 1999.) Since, no precise information was available on the efficacy of neem formulation against *A. solani* caused early blight in tomato.

Bio-inoculants viz. T. viride, T. harzianum, Bacillus subtilis, Pseudomonas fluorescence were reported efficient antagonists against A. solani earlier by many workers (Bais et al., 2019; Naik et al., 2020; Sudarshan et al., 2020, Verma et al., 2020; Mohamed et al., 2021). The species of Trichoderma viz. viride and

Choudhary et al.,

harzianum were reported as efficient antagonists against *Alternaria* spp (Pun *et al.*, 2020; Zin and Badaluddin, 2020). The fungistatic antifungal action exerted by the species of *Trichoderma* and against *A. solani* and other species of *Alternaria* may be attributed to their production of volatile and non-volatile substances, cell wall degrading enzymes (glucanases, Bl, 3 glucanase), the phenomenon of competition, lysis and antibiosis (Contreras-Cornejo *et al.*, 2015a, 2015b; Kubicek *et al.*, 2001; Kullnig *et al.*, 2000).

In conclusion, the study confirms that different neem formulation, neem oil (77.09%) at 1500 ppm and bioagents, *T harzianum* (71.15%) and *T. viride* (66.35%) are very effective against early blight caused by *Alternaria solani* of tomato.

Acknowledgement. The authors are sincerely thankful to the Dean College of Agriculture Jodhpur, Rajasthan for providing necessary facilities and guidance to conduct the different experiments.

Conflicts of Interest. The results furnished in this paper were from my own research and there were no any conflicts from other research scholars or scientists.

REFERENCES

- Adhikari, P., Yeonyee, O., & Panthee, D. R. (2017). Current status of Early blight resistance in tomato an update. *International Journal of Molecular Sciences*, 6(8): 2-22.
- Anonymous (2020). National Horticulture Board (3rd Advanced estimation) http://nhb.gov.in/statistics/State Level/2018.
- Bais, R.K., Ratan, V., Kumar, S., & Tiwari, A. (2019). Comparative analysis of various strategies for management of early blight of tomato incited by *Alternaria solani* (Ellis and Martin) Jones and Grout. *The Pharma Innovation Journal*, 8(12): 15-22.
- Bessadat, N., Benichou, S., Kihal, M., & Henni, D. E. (2014). Aggressiveness and morphological variability of small spore Alternaria species isolated from Algeria. Journal of Experimental Biology and Agriculture Sciences, 2: 266–278.
- Bhonde, S. B., Deshpande, S. G., & Sharma, R. N. (1999). In vitro evaluation on inhibitory nature of some Neem formulations against plant pathogenic fungi. *Hindustan antibiotics bulletin*, 41(1-4): 22-24.
- Chaudhary, R. F., Patel, R. L., Chaudhari, S. M., Pandey, S. K., & Brajesh, S. (2003). *In vitro* evaluation of different plant extracts against *Alternaria alternata* causing early blight of potato. *Journal of the Indian Potato Association*, 30(1/2): 141-142.
- Contreras-Cornejo, H. A., Macias-Rodriguez, L., Vergara, A. G., & Lopez-Bucio, J. (2015b). Trichoderma modulates stomatal aperture and leaf transpiration through an abscisic acid-dependent mechanism in Arabidopsis. Journal of Plant Growth Regulation, 34(2): 425-432.
- Contreras-Cornejo, Hexon Angel, Jesus Salvador Lopez-Bucio, Alejandro Mendez-Bravo, Lourdes Macías-Rodriguez, Maricela Ramos-Vega, Angel Arturo Guevara-Garcia, & Jose Lopez-Bucio. (2015a). Mitogen-activated protein kinase 6 and ethylene and auxin signaling pathways are involved in Arabidopsis root-system architecture alterations by *Trichoderma atroviride*." *Molecular Plant-Microbe Interactions*, 28(6): 701-710.

- Cook, R.J. (2000). Advances in plant health management in the 20th century. *Annual Review* of *Phytopathology* 38: 95–116.
- Datar, V. V., & Mayee, C. D. (1981). Assessment of losses in tomato yield due to early blight. *Indian Phytopathology*, 34: 191-195.
- Denis, C., & Webster, J. (1971). Antagonistic properties of species group of *Trichoderma*. Production of volatile antibiotics. *Transactions of the British Mycological Society*, 57: 41-48.
- Girish, K., & Shankara, B. S. (2008). Neem–a green treasure. *Electronic journal of Biology*, 4(3): 102-111.
 Golyshin, N. . (1990). *Plant Protection*, 11: 13-15.
- Guleria, S., & Kumar, A., (2006). Azadirachta indica leaf extract induces resistance in sesame against Alternaria leaf spot disease. *Journal of Cell and Molecular Biology*, 5(2): 81-86.
- Kale, P.N., & Kale, S. P. (1994). Bhajipala Utpadan (Vegetable production). Continental Publication, Co., Pune. Pp-29-30.
- Kemmitt, G. (2002). Early blight of potato and tomato. The Plant Health Instructor. DOI: https: //doi: 10.1094/PHI-I-2002-0809-01.
- Kota V. (2003). Biological management of postharvest fungal diseases of major fruits [M.Sc thesis]. India: University of Agricultural Sciences.
- Kubicek, C. P., Mach, R. L., Peterbauer, C. K., & Lorito, M. (2001). Trichoderma: from genes to biocontrol. Journal of Plant Pathology, pp. 11-23.
- Kullnig, C., Mach, R. L., Lorito, M., & Kubicek, C. P. (2000). Enzyme diffusion from *Trichoderma atroviride* (= *T. harzianum* P1) to *Rhizoctonia solani* is a prerequisite for triggering of Trichoderma ech42 gene expression before mycoparasitic contact. *Applied and Environmental Microbiology*, 66(5): 2232-2234.
- Kumar A. (2008). Studies on leaf blight of Chrysanthemum caused by *Alternaria alternata* (Fr.) Keissler [M.Sc thesis]. India: University of Agricultural Sciences.
- Mohamed, A. A., Salah, M. M., El-Dein, M. M. Z., EL-Hefny, M., Ali, H. M., Farraj, D. A. A., Hatamleh, A. A., Salem, M. Z., & Ashmawy, N. A. (2021). Ecofriendly Bioagents, Parthenocissus quinquefolia, and Plectranthus neochilus Extracts to Control the Early Blight Pathogen (*Alternaria solani*) in Tomato. *Agronomy*, 11(5): 1-17.
- Naik, S. C., Narute, T. K., Narute, T. T., & Khaire, P. B. (2020). *In-vitro* efficacy of biocontrol agents against *Alternaria solani* (Early Blight of Tomato). *Journal of Pharmacognosy and Phytochemistry*; 9(5): 550-552.
- Nene, Y. L., & Thapliyal P. N. (1993). Fungicides in Plant Disease Control. Oxford and IBH publishing company, New Delhi, 531.
- Peralta, I. E., Knapp, S., & Spooner, D. M. (2005). New species of wild tomatoes (*Solanum* section *Lycopersicon*: Solanaceae) from northern Peru. *Systematic Botany*, 30: 424–434.
- Pun, L. B., Chhetri, K., Pandey, A. & Poudel, R. (2020). In vitro Evaluation of Botanical Extracts, Chemical Fungicides and Trichoderma harzianum against Alternaria brassicicola causing Leafspot of Cabbage. Nepalese Horticulture, 14(1): 68-76.
- Saha, D., Dasgupta, S., & Saha, A. (2005). Antifungal activity of some plant extracts against fungal pathogens of tea (*Camellia sinensis*). *Pharmaceutical biology*, 43(1): 87-91.
- Sudarshan G. K., Nagaraj, M. S., Thammaiah N., Yogananada, S. B., Mallikarjuna Gowda, A. P., & Prasanna Kumar, M. K. (2020). *In vitro* Efficacy of

Choudhary et al.,

Biological Forum – An International Journal 13(4): 45-50(2021)

Fungicides and Bioagents against Early Blight of Tomato caused by Alternaria solani. International Journal of Current Microbiology and Applied Sciences, 9(9): 1490-1496.

- Vazquez, M. B., Barrera, V., & Bianchinotti, V. (2015). Molecular identification of three isolates of *Trichoderma harzianum* isolated from agricultural soils in Argentina, and their abilities to detoxify *in vitro* metsulfuron methyl. *Botany*, 93(11): 793-800.
- Vennila, S., Bhat, M. N., Kumari, D. A., Yadav, S. K., & Sharma, V. K. (2020). Effect of climatic variability and weather factors on development of tomato early blight in a hot semi-arid region of Southern India. *Indian Journal of Horticulture Science*, 77(2): 333-338.
- Verma, A., Shukla, A., & Singh, N. (2020). In vitro evaluation of botanicals, bio-agents & chemical fungicides against Alternaria solani (Ellis & Martin)

Sorauer causing early blight in tomato. Journal of Pharmacognosy and Phytochemistry, 9(5): 2377-2380.

- Vincent, J. M. (1947). Distortion of fungal hyphae in the presence of certain inhibitors. *Nature*, 159(4): 850-850.
- Wang, J., Li, J., Cao, J., & Jiang, W. (2010). Antifungal activities of neem (*Azadirachta indica*) seed kernel extracts on postharvest diseases in fruits. *African Journal of Microbiology Research*, 4(11): 1100-1104.
- Zafra, G., Moreno-Montaño, A., Absalón, Á. E., & Cortés-Espinosa, D. V. (2015). Degradation of polycyclic aromatic hydrocarbons in soil by a tolerant strain of *Trichoderma asperellum. Environmental Science and Pollution Research*, 22(2): 1034-1042.
- Zin, N. A., & Badaluddin, N. A. (2020). Biological functions of *Trichoderma spp.* for agriculture applications. *Annals of Agricultural Sciences*, 65(2): 168-178.

How to cite this article: Choudhary, A., Verma, J.R., Ram, D., Yadav, P. (2021). Management of Early Blight of Tomato through Neem Formulation and Bio-inoculants. *Biological Forum – An International Journal*, *13*(4): 45-50.